

By preliminary amendment, Applicants correct certain erroneous references to the sequence identifiers and add sequence identifiers where appropriate throughout the specification. These corrections and additions are shown in detail in the marked-up version of the affected paragraphs attached hereto as "Appendix - 1. "Revisions Made to Specification and Claims." Claims 7-21 and 34-37 have been canceled, and claims 3-6 and 24, 29, 30 and 32 have been amended. The changes to the amended claims are also shown in detail in Appendix - 1. A complete set of the pending claims, as presently amended, is attached as "Appendix - 2. Complete Set of Pending Claims, as Amended on September 17, 2001."

In the "Notice to File Missing Parts of Nonprovisional Application," the Initial Patent Examiner stated that the application is informal for the reason that excessive text is contained in the drawings as originally filed. In reply, substitute sheets of the drawings are submitted containing Figures 1-12 to replace the set of drawings filed on April 10, 2001. As required by the Initial Patent Examiner, the Figures have been revised to omit certain text so as to better comply with 37 CFR § 1.84. Text deleted from the Figures has been moved to the corresponding figure descriptions in the "Brief Description of the Drawings" section of the Specification, or has been moved to the "Detailed Description" section where the particular Figure is discussed, or has been deleted because it is considered duplicative of description in the Specification as originally filed. A marked-up copy of the Figures is attached as "Appendix - 3. Details of Drawing Revisions Submitted on September 17, 2001," showing the proposed changes by circling the new text in red ink and by striking out deleted text in red ink. If the consent of the Examiner is necessary for entry of these substitute sheets of drawings, Applicants respectfully request approval by the Examiner of the proposed corrections to the Figures and entry of the substitute sheets of drawings.

Also enclosed is a computer diskette containing the initial computer readable form (CRF) of the Sequence Listing and a printed copy of the same, along with a statement that the content of the CRF and the original printed copy attached to the Application as filed is the same, and that the CRF and the attached printed copy are the same, and that no new matter is introduced thereby.

Applicants respectfully submit that the amendments to the specification, claims and drawings presented herein are to correct typographical errors, to facilitate examination of the application by ensuring compliance with 37 CFR § 1.821-1.824 or 37 CFR § 1.75(c) or 37 CFR § 1.84 prior to the first examination on the merits, or to comply with specific requirements stated in the "Notice to File Missing Parts." No new matter is introduced by way of these amendments.

If any additional fees are required or if the fees submitted are in excess of that required, please charge or credit the appropriate fees to Conley, Rose & Tayon, P.C. Deposit Account No.

03-2769/1889/00401/CGM. In the event that an extension of time is necessary in order for this submission to be deemed timely filed, please consider this a petition therefor and charge the associated fee to Conley, Rose & Tayon, P.C. Deposit Account No. 03-2769/1889/00401/CGM.

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APPENDIX - 1

REVISIONS MADE TO REPLACEMENT PARAGRAPHS AND CLAIMS

[0005] The invention relates to a compound that competitively inhibits binding of CSP [SEQ ID NO:4] to *S. mutans* histidine kinase [SEQ ID NO:6]. The compound is preferably a peptide or an antibody. The compound is preferably a derivative of [SEQ ID NO:2] [SEQ ID NO:3], a fragment of [SEQ ID NO:2] [SEQ ID NO:3] or a derivative of a fragment of [SEQ ID NO:2] [SEQ ID NO:3].

[0006] Schematic layout of the arrangement of the genetic locus encoding the signal peptide precursor (ComC) [SEQ ID NO:1], the histidine kinase (ComD) [SEQ ID NO:5] and the response regulator (ComE) [SEQ ID NO:7]. Note that this arrangement is different from other loci in related streptococci for the following reasons: a) The comC gene [SEQ ID NO:1] is transcribed from its own unique promoter, unlike the genes thus far described in other streptococci that are arranged in an operon-like cluster with the comC/DE genes being transcribed from a single promoter.

[0007] b) The comC gene [SEQ ID NO:1] is separated by 148 nucleotides from the comD gene [SEQ ID NO:5].

[0008] Shows the nucleic acid molecule that is [SEQ ID NO:1] [SEQ ID NO:3]. In a preferred embodiment, the figure shows CSP (competence signal peptide [SEQ ID NO:3]). Nucleotide sequence of the locus. Figure 2 also shows histidine kinase [SEQ ID NO:5] sequences and response regulator [SEQ ID NO:7] sequences.

[0008.5] Figure 2A. *S. mutans* comC gene [SEQ ID NO:1]. Encodes a precursor to a signal peptide [SEQ ID NO:2].

[0008.6] Figure 2B. *S. mutans* CSP encoding sequence [SEQ ID NO:3]. Encodes a Competence Signal Peptide [SEQ ID NO:4].

[0008.7] Figure 2C. *S. mutans* comD gene [SEQ ID NO:5]. Encodes a protein that functions as a histidine kinase receptor [SEQ ID NO:6].

[0008.8] Figure 2D. *S. mutans* comE gene [SEQ ID NO:7]. Encodes a response regulator that activates transcription of a number of genes [SEQ ID NO:8].

[0009] Sequence of the deduced amino acid sequence of the signal peptide [SEQ ID NO:4], histidine kinase [SEQ ID NO:6], and response regulator [SEQ ID NO:8].

[0009.4] Figure 3A. *S. mutans* ComC protein (CSP Precursor) [SEQ ID NO:2].

[0009.5] Figure 3B. *S. mutans* ComD protein (Histidine Kinase) [SEQ ID NO:6].

[0009.6] Figure 3C. *S. mutans* ComE protein (Response Regulator) [SEQ ID NO:8].

[0010] The deduced amino acid sequence of the signal peptide precursor in various strains and its predicted cleavage site, following the glycine-glycine signal The original peptide is expressed as a 46 amino acid peptide that is cleaved after the glycine-glycine residues to generate an active signal peptide.

[0011] Shows the peptide that is [SEQ ID NO:2] **[SEQ ID NO:4]**. The synthetic signal peptide **[SEQ ID NO:16]** that is effective at inducing competence, biofilm formation and acid tolerance in *Streptococcus mutans*.

[0013] Table illustrating the effect of synthetic peptide on genetic competence in *S. mutans* cells. Induction of genetic transformation in *Streptococcus mutans* by synthetic competence stimulating peptide (SCSP).

[0015] ComCDE local region **[SEQ ID NO:23]**. The ComC (first highlighted region; nucleotide 101 to 241), ComD (second highlighted region; nucleotides 383 to 1708) and ComE (third highlighted region; nucleotides 1705 to 2457) proteins are highlighted.

[0016] The comX DNA sequence **[SEQ ID NO:22]**, protein sequence **[SEQ ID NO:23]**, and the comX gene local region **[SEQ ID NO:24]** with 100bp included both upstream and downstream (promoter is upstream).

[0016.4] Figure 10A. *S. mutans* comX gene [SEQ ID NO:22].

[0016.5] Figure 10B. *S. mutans* ComX protein [SEQ ID NO:25].

[0016.6] Figure 10C. *S. mutans* comX gene local region [SEQ ID NO:26].

[0017] The comA and comB nucleotide **[SEQ ID NO:25]** and **[SEQ ID NO:27]** and amino acid sequences **[SEQ ID NO:26]** and **[SEQ ID NO:28]**. ComA and ComB are the components of the CSP exporter.

[0017.5] Figure 11A. *S. mutans* comA gene [SEQ ID NO:27].

[0017.6] Figure 11B. *S. mutans* ComA protein [SEQ ID NO:28].

[0017.7] Figure 11C. *S. mutans* comB gene [SEQ ID NO:29].

[0017.8] Figure 11D. *S. mutans* ComB protein [SEQ ID NO:30].

[0018] Illustrates the effect of synthetic peptide on acid resistance tolerance in *S. mutans* comC deficient cells. Addition of synthetic signal peptide (CSP) [SEQ ID NO:16] into the culture of the comC mutant restored the ability of the mutant to survive a low pH challenge when compared to the parent strain NG8.

[0020] We have identified a genetic locus in *S. mutans* consisting of three genes that encode: 1) a peptide precursor **[SEQ ID NO:2]** that is processed during export into a secreted 21-amino acid peptide (CSP) **[SEQ ID NO:4]**; 2) a histidine kinase **[SEQ ID NO:6]** that acts as a cell surface receptor activated by the peptide; 3) a response regulator **[SEQ ID NO:8]** that activates a number of other genes involved in genetic competence, biofilm formation, and acid tolerance of *S. mutans*. These properties have been attributed to the bacterium's ability to cause dental caries. Inactivation of any of these three genes or impairment of interaction or activity of any of their encoded proteins will disrupt the bacterium's ability to take up foreign DNA, form biofilms, and tolerate acidic pH.

[0021] *Streptococcus mutans* is a resident of the biofilm environment of dental plaque, a matrix of bacteria and extracellular material that adheres to the tooth surface. Under appropriate environmental conditions populations of *S. mutans* and the pH of the surrounding plaque will drop. *S. mutans*, being among the most acid tolerant organisms residing in dental plaque, will increase its numbers in this acidic environment and eventually become a dominant member of the plaque community. This situation eventually leads to dissolution of the tooth enamel, resulting in the development of dental caries. We control the accumulation and acid tolerance of this bacterium to make it less able to cause caries. We accomplish this by using inhibitors of an extracellular signal peptide that promotes the expression of genes involved in *S. mutans* biofilm formation and acid tolerance. The invention includes compounds that inhibit the action of the peptide. These inhibitors can include peptides, antibodies, or other agents that specifically inhibit the activation of the histidine kinase and the family of genes activated as a result of the histidine kinase activation by the signal molecule. Inhibitors include: modified structures of the peptide where amino acids are removed from the N- and/or COOH terminal of the peptide and/or substitutions of internal amino acid residues. We delete, one, two to 5, 6 to 10 and 10 to 15 amino acids from the peptide (for example at either terminal) and measure competitive inhibition of signal peptide binding to histidine

[0022] kinase (1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 or more amino acids are deleted and inhibition measured). Inhibitors also include antibodies raised against the 21-amino acid CSP **[SEQ ID NO:4]** alone or coupled to a larger molecule to increase immunogenicity.

[0024] In addition to identifying the genes encoding this signaling/sensing system, we have identified and chemically synthesized a 21-amino acid peptide **[SEQ ID NO:16]** that promotes biofilm formation and acid tolerance of *S. mutans*. A survey of the literature and genome databases reveals that genes similar to this signal-receptor system are present in most Gram-

positive bacteria, and therefore an inhibitor, or family of related inhibitors may be effective at inhibiting biofilm formation among a large group of bacteria.

[0025] The invention treats or prevents dental caries by addition of compounds that inhibit the stimulatory action of the 21-amino acid peptide [SEQ ID NO:4] on biofilm formation and acid tolerance of *S. mutans*. This is accomplished by delivery of these compounds to the biofilm and/or to incorporate these inhibitors into materials to control growth on surfaces. This includes delivery by topical application, alone or in combination with other compounds including toothpaste, mouthwash, food or food additives.

[0027] The invention includes an isolated CSP from *S. mutans*. The invention also includes a recombinant isolated CSP [SEQ ID NO:4] peptide produced by a cell including a nucleic acid molecule encoding CSP [SEQ ID NO:3] operably linked to a promoter. The invention also includes an isolated nucleic acid molecule encoding a CSP [SEQ ID NO:3]. The peptide we work with is preferably chemically synthesized [SEQ ID NO:16].

[0028] The invention includes CSP-encoding nucleic acid molecules [SEQ ID NO:3] and molecules having sequence identity or which hybridize to the CSP-encoding sequence and which encode a peptide having CSP activity (preferred percentages for sequence identity are described below) as well as vectors including these molecules. The invention also includes CSP [SEQ ID NO:4] or peptides having sequence identity (preferred percentages described below) or which have CSP activity. The nucleic acid molecules and peptides of the invention may be from *S. mutans* and they may be isolated from a native source, synthetic or recombinant. The invention includes CSP [SEQ ID NO:4] or peptides having sequence identity, which have CSP activity, as prepared by the processes described in this application.

[0029] The invention includes an isolated HK [SEQ ID NO:6] from *S. mutans*. The invention also includes a recombinant isolated HK polypeptide produced by a cell including a nucleic acid molecule encoding HK [SEQ ID NO:5] operably linked to a promoter. The invention also includes an isolated nucleic acid molecule encoding a HK polypeptide [SEQ ID NO:6].

[0030] The invention includes HK-encoding nucleic acid molecules and molecules having sequence identity or which hybridize to the HK-encoding sequence [SEQ ID NO:5] and which encode a protein having HK activity (preferred percentages for sequence identity are described below) as well as vectors including these molecules. The invention also includes HK [SEQ ID NO:4] or polypeptides having sequence identity (preferred percentages described below) or which have HK activity. The nucleic acid molecules and polypeptides of the invention may be

from *S. mutans* and they may be isolated from a native source, synthetic or recombinant. The invention includes HK [SEQ ID NO:4] or polypeptides having sequence identity, which have HK activity, as prepared by the processes described in this application.

[0031] The invention includes an isolated RR [SEQ ID NO:6] from *S. mutans*. The invention also includes a recombinant isolated RR [SEQ ID NO:6] polypeptide produced by a cell including a nucleic acid molecule encoding RR [SEQ ID NO:6] operably linked to a promoter. The invention also includes an isolated nucleic acid molecule encoding a RR polypeptide.

[0032] The invention includes RR-encoding nucleic acid molecules and molecules having sequence identity or which hybridize to the RR-encoding sequence [SEQ ID NO:5] and which encode a polypeptide having RR activity (preferred percentages for sequence identity are described below) as well as vectors including these molecules. The invention also includes RR [SEQ ID NO:6] or polypeptides having sequence identity (preferred percentages described below) or which have RR activity. The nucleic acid molecules and polypeptides of the invention may be from *S. mutans* and they may be isolated from a native source, synthetic or recombinant. The invention includes RR [SEQ ID NO:6] or polypeptides having sequence identity, which have RR activity, as prepared by the processes described in this application.

[0033] The comA and comB nucleotide [SEQ ID NO:27 and SEQ ID NO:29] and amino acid sequences [SEQ ID NO:28 and SEQ ID NO:30] are also aspects of the invention. ComA and ComB are components of the CSP exporter. The discussion of variants, sequence identity etc. for CSP, HK, RR applies to both the full sequences shown in the figures as well as bracketed portions of sequences (coding regions). The peptides and polypeptides may be natural, recombinantly produced or synthetic.

[0034] The invention includes nucleic acid molecules that are functional equivalents of all or part of the CSP sequence in [SEQ ID NO:1] [SEQ ID NO:3]. (A nucleic acid molecule may also be referred to as a DNA sequence or nucleotide sequence in this application. All these terms have the same meaning as nucleic acid molecule). Functionally equivalent nucleic acid molecules are DNA and RNA (such as genomic DNA, complementary DNA, synthetic DNA, and messenger RNA molecules) that encode peptides having the same or similar CSP activity as the CSP peptide shown in [SEQ ID NO:2] [SEQ ID NO:4]. Functionally equivalent nucleic acid molecules can encode peptides that contain a region having sequence identity to a region of a CSP peptide [SEQ ID NO:4] or more preferably to the entire CSP peptide. Identity is calculated according to methods known in the art. The ClustalW program (preferably using default parameters) [Thompson, JD et al., Nucleic Acid Res. 22:4673-4680.], described below, is most preferred. For example, if a nucleic acid molecule (called "Sequence A") has 90% identity to a

portion of the nucleic acid molecule in [SEQ ID NO:1] [SEQ ID NO:3], then Sequence A will preferably be identical to the referenced portion of the nucleic acid molecule in [SEQ ID NO:1] [SEQ ID NO:3], except that Sequence A may include up to 10 point mutations, such as substitutions with other nucleotides, per each 100 nucleotides of the referenced portion of the nucleic acid molecule in [SEQ ID NO:1] [SEQ ID NO:3]. Mutations described in this application preferably do not disrupt the reading frame of the coding sequence. Nucleic acid molecules functionally equivalent to the CSP sequences can occur in a variety of forms as described below.

[0035] Nucleic acid molecules may encode conservative amino acid changes in CSP peptide [SEQ ID NO:4]. The invention includes functionally equivalent nucleic acid molecules that encode conservative amino acid changes within a CSP amino acid sequence and produce silent amino acid changes in CSP.

[0036] Nucleic acid molecules may encode non-conservative amino acid substitutions, additions or deletions in CSP peptide. The invention includes functionally equivalent nucleic acid molecules that make non-conservative amino acid changes within the CSP amino acid sequence in [SEQ ID NO:2] [SEQ ID NO:4]. Functionally equivalent nucleic acid molecules include DNA and RNA that encode peptides, peptides and proteins having non-conservative amino acid substitutions (preferably substitution of a chemically similar amino acid), additions, or deletions but which also retain the same or similar CSP activity as the CSP peptide shown in [SEQ ID NO:2] [SEQ ID NO:4]. The DNA or RNA can encode fragments or variants of CSP. Fragments are useful as immunogens and in immunogenic compositions (U.S. Patent No. 5,837,472). The CSP or CSP-like activity of such fragments and variants is identified by assays as described below. Fragments and variants of CSP encompassed by the present invention should preferably have at least about 40%, 60%, 80% or 95% sequence identity to the naturally occurring CSP nucleic acid molecule, or a region of the sequence, such as the coding sequence or one of the conserved domains of the nucleic acid molecule, without being identical to the sequence in [SEQ ID NO:1] [SEQ ID NO:3]. Sequence identity is preferably measured with the ClustalW program (preferably using default parameters) (Thompson, JD et al., Nucleic Acid Res. 22:4673-4680)

[0037] Nucleic acid molecules functionally equivalent to the CSP nucleic acid molecule in [SEQ ID NO:1] [SEQ ID NO:3] will be apparent from the following description. For example, the sequence shown in [SEQ ID NO:1] [SEQ ID NO:3] may have its length altered by natural or artificial mutations such as partial nucleotide insertion or deletion, so that when the entire length of the coding sequence within [SEQ ID NO:1] [SEQ ID NO:3], is taken as 100%, the functional

equivalent nucleic acid molecule preferably has a length of about 60-120% thereof, more preferably about 80-110% thereof. Fragments may be less than 60%.

[0038] Nucleic acid molecules containing partial (usually 80% or less, preferably 60% or less, more preferably 40% or less of the entire length) natural or artificial mutations so that some codons in these sequences code for different amino acids, but wherein the resulting peptide retains the same or similar CSP activity as that of a naturally occurring CSP peptide [SEQ ID NO:4]. The mutated DNAs created in this manner should preferably encode a peptide having at least about 40%, preferably at least about 60%, at least about 80%, and more preferably at least about 90% or 95% sequence identity to the amino acid sequence of the CSP peptide in [SEQ ID NO:2] [SEQ ID NO:4]. The ClustalW program preferably assesses sequence identity.

[0039] Since the genetic code is degenerate, the nucleic acid sequence in [SEQ ID NO:1] [SEQ ID NO:3] is not the only sequence which may code for a peptide having CSP activity. This invention includes nucleic acid molecules that have the same essential genetic information as the nucleic acid molecule described in [SEQ ID NO:1] [SEQ ID NO:3]. Nucleic acid molecules (including RNA) having one or more nucleic acid changes compared to the sequences described in this application and which result in production of a peptide shown in [SEQ ID NO:2] [SEQ ID NO:4] are within the scope of the invention.

[0040] Other functional equivalent forms of CSP-encoding nucleic acids can be isolated using conventional DNA-DNA or DNA-RNA hybridization techniques. Thus, the present invention also includes nucleic acid molecules that hybridize to one or more of the sequences in [SEQ ID NO:1] [SEQ ID NO:3] or its complementary sequence, and that encode expression for peptides, peptides and proteins exhibiting the same or similar activity as that of the CSP peptide produced by the DNA in [SEQ ID NO:1] [SEQ ID NO:3] or its variants. Such nucleic acid molecules preferably hybridize to the sequence in [SEQ ID NO:1] [SEQ ID NO:3] under moderate to high stringency conditions (see Sambrook et al. Molecular Cloning: A Laboratory Manual, Most Recent Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). High stringency washes have low salt (preferably about 0.2% SSC), and low stringency washes have high salt (preferably about 2% SSC). A temperature of about 37 °C or about 42 °C is considered low stringency, and a temperature of about 50-65 °C is high stringency. The invention also includes a method of identifying nucleic acid molecules encoding a CSP activator peptide (preferably a mammalian peptide), including contacting a sample containing nucleic acid molecules including all or part of [SEQ ID NO:1] [SEQ ID NO:3] (preferably at least about 15 or 20 nucleotides of [SEQ ID NO:1] [SEQ ID NO:3]) under moderate or high stringency hybridization conditions and identifying nucleic acid molecules which hybridize to the nucleic

acid molecules including all or part of [SEQ ID NO:1] [SEQ ID NO:3].) Similar methods are described in U.S. Patent No. 5,851,788, which is incorporated by reference in its entirety.

[0041] The invention also includes methods of using all or part of the nucleic acid molecules which hybridize to all or part of [SEQ ID NO:1] [SEQ ID NO:3], for example as probes or in assays to identify antagonists or inhibitors of the peptides produced by the nucleic acid molecules (described below). The invention also includes methods of using nucleic acid molecules having sequence identity to the CSP nucleic acid molecule [SEQ ID NO:3] (as described below) in similar methods.

[0042] The invention also includes a nucleic acid molecule detection kit including, preferably in a suitable container means or attached to a surface, a nucleic acid molecule of the invention encoding CSP [SEQ ID NO:4] or a peptide having CSP activity and a detection reagent (such as a detectable label). Other variants of kits will be apparent from this description and teachings in patents such as U.S. Patent Nos. 5,837,472 and 5,801,233, which are incorporated by reference in their entirety.

[0043] A nucleic acid molecule described above is considered to have a function substantially equivalent to the CSP nucleic acid molecules [SEQ ID NO:3] of the present invention if the peptide [SEQ ID NO:4] produced by the nucleic acid molecule has CSP activity. A peptide has CSP activity if it can stimulate genetic competence and acid tolerance in *S. mutans*.

Activation of the HK [SEQ ID NO:6]/RR [SEQ ID NO:8] is shown where a peptide is capable of stimulating the uptake and incorporation of foreign DNA. We describe below how the activity of these peptide-mediated processes can be measured by determining the efficiency of plasmid uptake, which is a measure of genetic competence. Since the ability to transport and incorporate foreign DNA relies on activation of the HK [SEQ ID NO:6]/RR [SEQ ID NO:8] and subsequent genes activated by the signal cascade initiated by the signal peptide, measurement of the conferment of erythromycin resistance by cells exposed to the peptide and plasmid DNA conferring erythromycin resistance indicates its level of function. Conversely if an inhibitor is capable of interfering with the action of the peptide the competence assay will indicate this by a corresponding decrease in the number of cells that acquire erythromycin resistance as described in the assays below (assays of genetic competence and assay of transformation of biofilms). Activation of the HK [SEQ ID NO:6]/RR [SEQ ID NO:8] is also shown where a peptide is capable of stimulating an acid tolerance response. We describe below how the activity of these peptide-mediated processes can be measured by determining the survival rate of cells in acidic pH conditions. Since the ability to survive exposure to acidic pH depends on the activation of the HK/RR and subsequent genes activated by the signal peptide, measurement of the survival of *S. mutans* in low pH conditions indicates the level of function of

the signal peptide. Conversely, if an inhibitor is capable of interfering with the signal peptide sensing system the assay for acid adaptation will indicate this by a corresponding decrease in the survival rate of cells grown in acidic pH conditions as described in the assay below (assay of acid adaptation).

[0049] Another embodiment of the invention relates to a method of transfecting a cell with a nucleic acid molecule of the invention, inserted in an expression vector to produce a cell expressing the CSP peptide [SEQ ID NO:4] or other peptide of the invention. The invention also relates to a method of expressing the peptides of the invention in a cell. A preferred process would include culturing a cell including a recombinant DNA vector including a nucleic acid molecule encoding CSP [SEQ ID NO:3] (or another nucleic acid molecule of the invention) in a culture medium so that the peptide is expressed. The process preferably further includes recovering the peptide from the cells or culture medium.

[0050] The invention also includes oligonucleotide probes made from the cloned CSP nucleic acid molecules described in this application or other nucleic acid molecules of the invention (see Materials and Methods section). The probes may be 15 to 20 nucleotides in length. A preferred probe is at least 15 nucleotides of CSP in [SEQ ID NO:1] [SEQ ID NO:3]. The invention also includes at least 15 consecutive nucleotides of [SEQ ID NO:1] [SEQ ID NO:3]. The probes are useful to identify nucleic acids encoding CSP peptides as well as peptides functionally equivalent to CSP. The oligonucleotide probes are capable of hybridizing to the sequence shown in [SEQ ID NO:1] [SEQ ID NO:3] under stringent hybridization conditions. A nucleic acid molecule encoding a peptide of the invention may be isolated from other organisms by screening a library under moderate to high stringency hybridization conditions with a labeled probe. The activity of the peptide encoded by the nucleic acid molecule is assessed by cloning and expression of the DNA. After the expression product is isolated, the peptide is assayed for CSP activity as described in this application.

[0051] Functionally equivalent CSP nucleic acid molecules from other cells, or equivalent CSP-encoding cDNAs or synthetic DNAs, can also be isolated by amplification using Polymerase Chain Reaction (PCR) methods. Oligonucleotide primers, such as degenerate primers, based on [SEQ ID NO:1] [SEQ ID NO:3] can be prepared and used with PCR and reverse transcriptase (E. S. Kawasaki (1990), In Innis et al., Eds., PCR Protocols, Academic Press, San Diego, Chapter 3, p. 21) to amplify functional equivalent DNAs from genomic or cDNA libraries of other organisms. The oligonucleotides can also be used as probes to screen cDNA libraries.

[0052] The present invention includes not only the peptides encoded by the sequences of the invention, but also functionally equivalent peptides, peptides and proteins that exhibit the same or similar CSP peptide activity. A peptide is considered to possess a function substantially equivalent to that of the CSP peptide [SEQ ID NO:4] if it has CSP activity. CSP activity means that it is able to confer genetic competence to *S. mutans*, as measured by an increased ability to incorporate and express foreign genetic material, when added to cells as described in the assay of genetic competence below. CSP activity also means that the peptide is able to confer an acid tolerance response in *S. mutans* as measured by an increase in cell survival under acidic pH conditions when added to cells as described in the assay for acid adaptation below. Functionally equivalent peptides, peptides and proteins include peptides, peptides and proteins that have the same or similar protein activity as CSP when assayed, i.e. they are able to stimulate genetic competence and low pH tolerance (the ability to withstand acid challenges of pH 3.5 –pH 3.0 for up to 3 hours) in *S. mutans*. A peptide has CSP activity if it is capable of increasing the frequency of uptake and expression of foreign DNA as described in the following assay for genetic competence and if the peptide can promote an acid tolerance response as described in the assay for acid adaptation.

[0053] Identity refers to the similarity of two peptides or proteins that are aligned so that the highest order match is obtained. Identity is calculated according to methods known in the art, such as the ClustalW program. For example, if a peptide (called "Sequence A") has 90% identity to a portion of the peptide in [SEQ ID NO:2] [SEQ ID NO:16], then Sequence A will be identical to the referenced portion of the peptide in [SEQ ID NO:2] [SEQ ID NO:16], except that Sequence A may include up to 1 point mutations, such as substitutions with other amino acids, per each 10 amino acids of the referenced portion of the peptide in [SEQ ID NO:2] [SEQ ID NO:16]. Peptides, peptides and proteins functional equivalent to the CSP peptides can occur in a variety of forms as described below.

[0054] Peptides biologically equivalent in function to CSP peptide include amino acid sequences containing amino acid changes in the CSP sequence [SEQ ID NO:4]. The functional equivalent peptides have at least about 40% sequence identity, preferably at least about 60%, at least about 75%, at least about 80%, at least about 90% or at least about 95% sequence identity, to the natural CSP peptide [SEQ ID NO:4] or a corresponding region. The ClustalW program preferably determines sequence identity. Most preferably, 1, 2, 3, 4, 5, 5-10, 10-15 amino acids are modified.

[0058] The variants preferably retain the same or similar CSP activity as the naturally occurring CSP [SEQ ID NO:4]. The CSP activity of such variants can be assayed by techniques described in this application and known in the art.

[0059] Variants produced by combinations of the techniques described above but which retain the same or similar CSP activity as naturally occurring CSP [SEQ ID NO:4] are also included in the invention (for example, combinations of amino acid additions, and substitutions).

[0062] As well, probes and antibodies for a histidine kinase [SEQ ID NO:5 and SEQ ID NO:6], response regulator [SEQ ID NO:7 and SEQ ID NO:8] comA [SEQ ID NO:27 and SEQ ID NO:28] or comB [SEQ ID NO:29 and SEQ ID NO:30] may be prepared using the description in this application and techniques known in the art. The description for preparation of CSP variants and mutants is also applicable to the histidine kinase [SEQ ID NO:5 and SEQ ID NO:6], response regulator [SEQ ID NO:7 and SEQ ID NO:8] or comA [SEQ ID NO:27 and SEQ ID NO:28] and comB [SEQ ID NO:29 and SEQ ID NO:30] of the invention. The invention also includes fragments of HK [SEQ ID NO:5 and SEQ ID NO:6] having HK activity, fragments of RR [SEQ ID NO:7 and SEQ ID NO:8] having RR activity and fragments of comA [SEQ ID NO:27 and SEQ ID NO:28] or comB [SEQ ID NO:29 and SEQ ID NO:30] having activity.

[0063] The activity of the CSP peptide [SEQ ID NO:4] may be varied by carrying out selective site-directed mutagenesis. We characterize the binding domain and other critical amino acid residues in the peptide that are candidates for mutation, insertion and/or deletion. Sequence variants may be synthesized. A DNA plasmid or expression vector containing the CSP nucleic acid molecule [SEQ ID NO:3] or a nucleic acid molecule having sequence identity may be used for these studies using the U.S.E. (Unique site elimination) mutagenesis kit from Pharmacia Biotech or other mutagenesis kits that are commercially available, or using PCR. Once the mutation is created and confirmed by DNA sequence analysis, the mutant peptide is expressed using an expression system and its activity is monitored. This approach is useful to identify CSP inhibitors. All these modifications of the CSP DNA sequences [SEQ ID NO:3] presented in this application and the peptides produced by the modified sequences are encompassed by the present invention.

[0064] The CSP inhibitors are also useful when combined with a carrier in a pharmaceutical composition. The compositions are useful when administered in methods of medical treatment or prophylaxis of a disease, disorder or abnormal physical state caused by *S. mutans*. The invention also includes methods of medical treatment of a disease, disorder or abnormal

physical state characterized by excessive *S. mutans* or levels or activity of CSP peptide **[SEQ ID NO:4]**, for example by administering a pharmaceutical composition including a carrier and a CSP inhibitor. Caries is one example of a disease, which can be treated or prevented by antagonizing CSP **[SEQ ID NO:4]**.

[0066] CSP activity could be blocked by antisense mRNA or by inhibiting the activity of the exporter that secretes it from the cell. We have the sequence of these exporters. There are two copies of the genes (comAB) **[SEQ ID NO:27 and SEQ ID NO:29]** that are involved in export.

[0067] Nucleic acid molecules (antisense inhibitors of CSP) **[SEQ ID NO:3]** and competitive inhibitors of CSP **[SEQ ID NO:4]** may be introduced into cells using *in vivo* delivery vehicles such as liposomes. They may also be introduced into these cells using physical techniques such as microinjection and electroporation or chemical methods such as coprecipitation or using liposomes.

[0070] Antibodies directed against the CSP **[SEQ ID NO:4 or SEQ ID NO:16]** would provide protection against caries. Antibodies may be manufactured as described below. Alternatively, a peptide of the invention **[SEQ ID NO:4 or SEQ ID NO:16]** or a fragment thereof may be used with a carrier to make a vaccine. The peptide or fragment may also be conjugated to another molecule to increase its antigenicity. Antibodies can also be coupled to the peptide (Brady, L.J. et al., "Monoclonal Antibody-Mediated Modulation of the Humoral Immune Response against Mucosally Applied *Streptococcus mutans*" (in press). In order to enhance the immune response the peptide can be coupled to KLH, ovalbumin, or thyroglobulin prior to immunization. The vaccine composition will trigger the mammal's immune system to produce antibodies. The invention includes vaccine compositions and methods of vaccinating a mammal, preferably a human, against dental caries by administering to the mammal an effective amount of a vaccine composition. Techniques for preparing and using vaccines are known in the art. To prepare the vaccine, the peptide, or a fragment of the peptide, may be mixed with other antigens (of different immunogenicity), a vehicle or an excipient. Examples of peptide vaccines are found in U.S. Patent Nos. 5,679,352, 5,194,254 and 4,950,480. Techniques for preparing vaccines involving site-directed mutagenesis are described in U.S. Patent Nos. 5,714,372, 5,543,302, 5,433,945, 5,358,868, 5,332,583, 5,244,657, 5,221,618, 5,147,643, 5,085,862 and 5,073,494. Vaccines may be administered by known techniques, such as topical or parenteral administration. Vast changes are taking place in vaccinology consequent to the introduction of new technologies. Acellular purified fractions devoid of side effects, non-pathogenic but immunogenic mutants, recombinant technology, conjugated vaccines, combination vaccines (to limit the number of injections). Vaccine delivery systems can deliver multiple doses of the vaccine at a single

contact point. A genetically engineered oral vaccine is useful to impart better and longer duration of immunity. Oral vaccines are useful. The nose as a route for immunization is also useful. DNA alone can constitute the vaccines, inducing both humoral and cell-mediated immune responses. Live recombinant vaccines are also useful. Potent adjuvants add to the efficacy of the vaccines. One can also 'humanize' mouse monoclonals by genetic engineering and express these efficiently in plants. These recombinant antibodies are opening out an era of highly specific and safe therapeutic interventions. An advantage of preformed antibodies directed at a defined target and given in adequate amounts is the certainty of efficacy in every recipient, in contrast to vaccines, where the quality and quantum of immune response varies from individual to individual. For example, nasal immunization may be done as described in C. Jespersgaard et al. "Protective Immunity against *Streptococcus mutans* Infection in Mice after Intranasal Immunization with the Glucan-Binding Region of *S. mutans* Glucosyltransferase" Infection and Immunity, December 1999, p. 6543-6549, Vol. 67, No. 12. Vaccine compositions may comprise solid or liquid formulations such as gels, sprays, inhalants, tablets, toothpastes, mouthwashes or chewing gum.

[0072] Inhibitors are preferably directed towards CSP SEQ ID NO:4 or SEQ ID NO:16 to block *S. mutans* competence, low pH tolerance and biofilm formation.

[0073] A method of identifying a compound which reduces the interaction of CSP SEQ ID NO:4 or SEQ ID NO:16 with HK SEQ ID NO:6, can include: contacting (i) CSP SEQ ID NO:4 or SEQ ID NO:16 with (ii) HK SEQ ID NO:6, a CSP-binding fragment of HK SEQ ID NO:6 or a derivative of either of the foregoing in the presence of the compound; and b) determining whether the interaction between (i) and (ii) is reduced, thereby indicating that the compound reduces the interaction of CSP SEQ ID NO:4 or SEQ ID NO:16 and HK SEQ ID NO:6. A CSP inhibitor (caries treating or preventing compound) inhibits the interaction between (i) and (ii). By way of example, one can screen a synthetic peptide library. One could also screen small non-peptide organic molecules.

[0074] In one embodiment, the invention includes an assay for evaluating whether test compounds are capable of acting as agonists or antagonists for CSP SEQ ID NO:4, or a peptide having CSP functional activity, including culturing cells containing DNA which expresses CSP SEQ ID NO:3, or a peptide having CSP activity so that the culturing is carried out in the presence of at least one compound whose ability to modulate CSP activity is sought to be determined and thereafter monitoring the cells for either an increase or decrease in the level of CSP SEQ ID NO:4 or SEQ ID NO:16 or CSP activity. Other assays (as well as variations of

the above assay) will be apparent from the description of this invention and techniques such as those disclosed in U.S. Patent No. 5,851,788, 5,736,337 and 5,767,075 which are incorporated by reference in their entirety. For example, the test compound levels may be either fixed or variable.

[0075] The CSP [SEQ ID NO:4 or SEQ ID NO:16] peptide is also useful as an antigen for the preparation of antibodies that can be used to purify or detect other CSP-like peptides. Antibodies may also block CSP [SEQ ID NO:4] binding to HK [SEQ ID NO:6]. Antibodies are preferably targeted to the entire CSP [SEQ ID NO:4] sequence. The CSP peptide [SEQ ID NO:4 or SEQ ID NO:16] may be conjugated to other compounds, in order to increase immunogenicity.

[0076] We generate polyclonal antibodies against the CSP [SEQ ID NO:4 or SEQ ID NO:16], which is a unique sequence. Monoclonal and polyclonal antibodies are prepared according to the description in this application and techniques known in the art. For examples of methods of preparation and uses of monoclonal antibodies, see U.S. Patent Nos. 5,688,681, 5,688,657, 5,683,693, 5,667,781, 5,665,356, 5,591,628, 5,510,241, 5,503,987, 5,501,988, 5,500,345 and 5,496,705, which are incorporated by reference in their entirety. Examples of the preparation and uses of polyclonal antibodies are disclosed in U.S. Patent Nos. 5,512,282, 4,828,985, 5,225,331 and 5,124,147 which are incorporated by reference in their entirety. Antibodies recognizing CSP [SEQ ID NO:4 or SEQ ID NO:16] can be employed to screen organisms or tissues containing CSP peptide [SEQ ID NO:4] or CSP-like peptides. The antibodies are also valuable for immuno-purification of CSP [SEQ ID NO:4] or CSP-like peptides from crude extracts.

[0077] An antibody (preferably the antibody described above) may be used to detect CSP [SEQ ID NO:4] or a similar peptide, for example, by contacting a biological sample with the antibody under conditions allowing the formation of an immunological complex between the antibody and a peptide recognized by the antibody and detecting the presence or absence of the immunological complex whereby the presence of CSP [SEQ ID NO:4] or a similar peptide is detected in the sample. The invention also includes compositions preferably including the antibody, a medium suitable for the formation of an immunological complex between the antibody and a peptide recognized by the antibody and a reagent capable of detecting the immunological complex to ascertain the presence of CSP [SEQ ID NO:4] or a similar peptide. The invention also includes a kit for the *in vitro* detection of the presence or absence of CSP [SEQ ID NO:4] or a similar peptide in a biological sample, wherein the kit preferably includes an antibody, a medium suitable for the formation of an immunological complex between the antibody and a peptide recognized by the antibody and a reagent capable of detecting the

immunological complex to ascertain the presence of CSP [SEQ ID NO:4] or a similar peptide in a biological sample. Further background on the use of antibodies is provided, for example in U.S. Patent Nos. 5,695,931 and 5,837,472, which are incorporated by reference in their entirety.

[0078] The ability of the peptide to activate the HK [SEQ ID NO:6] and RR [SEQ ID NO:8] and the subsequent genes involved in the conferral of the properties of genetic competence, acid-tolerance and biofilm formation can be determined by measuring the efficiency of uptake and expression of DNA (preferably plasmid DNA) in *S. mutans* when exposed to signal peptide and/or inhibitor. Two methods modified based on the protocols described by Perry *et al.* Infect Immun, 41:722-727 and Lindler and Macrina J Bacteriol, 166:658-665 are used to assay genetic competence. The method involves adding DNA and CSP [SEQ ID NO:3] (preferably plasmid DNA) to a *S. mutans* culture (or culture of a bacteria expressing CSP [SEQ ID NO:3] or a variant thereof). The rate of transformation is then determined. *S. mutans* is preferably grown in THYE plus 5% horse serum (THYE-HS). After 2-hr incubation, 1 µg/ml plasmid DNA or 10 µg/ml of chromosomal DNA is added to the culture. To assay induction of competence, competence signal peptide, (SCSP) [SEQ ID NO:16] is then added to the cultures, incubation continued for 30 minutes with a final concentration of 500 ng/ml of SCSP added to each sample. After the 30-minute incubation equal amounts of DNA is added to each well (1 µg/ml plasmid or 10 µg/ml of chromosomal DNA) and incubation continued for another 2 hrs.. Cell dilutions were immediately spread on THYE agar plates plus appropriate antibiotics. Transformation frequency was expressed as the number of transformants (antibiotic resistant cells) per number of viable recipients. This is determined by comparing the number of cells able to grow in the presence of antibiotic (conferred by the applied plasmid or chromosomal DNA) relative to the total number of cells present (i.e., that grow in the absence of antibiotic). A higher value indicates a higher rate of transformation and thus is reflective of a stimulatory effect by the peptide. Consequently, addition of a molecule that successfully acts as an inhibitor results in a lower ratio of transformants/recipients, indicating that the inhibitor is effective at blocking activity of the CSP [SEQ ID NO:4]. CSP deficient cells [SEQ ID NO:3 or SEQ ID NO:4] may also be used in a variation of these assays. One can identify compounds that inhibit CSP [SEQ ID NO:4] or variants thereof by adding a test compound to the mixture to determine if the rate of transformation is decreased by the addition of the test compound.

[0079] The activity of the system can also be measured by an *in vitro* assay that relies on the measurement of marker protein expression (such as green fluorescent protein (GFP)) via expression from a fusion to a promoter controlled by the signal cascade initiated by CSP [SEQ ID NO:4]/HK [SEQ ID NO:6]/RR [SEQ ID NO:8]. One such promoter occurs immediately 5' proximal to the *S. mutans* comX gene. *S. mutans* cells grown in microtiter wells are exposed to

the CSP [SEQ ID NO:4] and/or inhibitor and the level of fluorescence of the comX::GFP strain is measured to give a quantitative measure of CSP [SEQ ID NO:4] stimulation (and conversely inhibitor activity). One can identify compounds that inhibit CSP [SEQ ID NO:4] or variants thereof by adding a test compound to the mixture to determine if the quantitative measure of CSP [SEQ ID NO:4] stimulation is decreased by the addition of the test compound.

[0080] The ability of CSP [SEQ ID NO:4] to promote acid resistance tolerance is determined by measuring the cell survival rate of *S. mutans* when exposed to acidic pH. In one example, *S. mutans* are first grown in batch culture to assay acid tolerance response in 'standard' log- and stationary-phase cells by using a modification of methods described previously by Svensäter *et al.* *Oral Microbiol. Immunol.*, 12:266-73. Mid-log-phase cells are obtained by transferring one volume of overnight culture into nine volumes (1:10) of fresh TYG medium (pH 7.5) and incubated at 37°C with 5% CO₂ for 2 hours. These cells are then collected by centrifugation at 8,000 x g for 10 min and resuspended in 2 ml of fresh TYG (pH 5.5) at various cell densities as determined by O.D₆₀₀. The cells are induced for acid adaptation by incubation at pH 5.5 for 2 h at 37°C with 5% CO₂. The adapted log-phase cells are then exposed to the killing pH. Killing pH is pre-determined by incubating unadapted, mid-log phase cells in TYG medium at pH values from 6.0 to 2.0. Stationary-phase cells are prepared by re-suspending late-log phase cells in TY medium (tryptone-yeast extract) without glucose. The culture is incubated at 37°C for 2 h to allow the cells to fully enter into stationary phase. Induction of acid adaptation in stationary-phase cells follows a similar procedure to that for log-phase cells. Adaptation of both log- and stationary-phase cells to acidic pH is determined by measuring the ability of bacterial cells to survive a killing pH for 3 h. Acid killing is initiated by resuspending cells in the same volume of fresh TYG (pH 3.5) and an aliquot of cell suspension is taken immediately from each sample to determine total viable cell number at zero time. The cells are then incubated for 3 h at 37°C with 5% CO₂ and an aliquot of sample is taken to determine survival rate by viable cell counts. Addition of a molecule that successfully acts as an inhibitor results in a decrease in the acid resistance tolerance of *S. mutans* resulting in a corresponding decrease in cell survival indicating that the inhibitor is effective at blocking activity of CSP [SEQ ID NO:4]. CSP [SEQ ID NO:3 or SEQ ID NO:4] deficient cells may also be used in a variation of these assays wherein addition of the signal peptide can complement the acid-adaptation-defective phenotype of a comC [SEQ ID NO:1 or SEQ ID NO:2] deficient cell. One can identify compounds that inhibit CSP [SEQ ID NO:3 or SEQ ID NO:4] or variants thereof by adding a test compound to the mixture to determine if the survival rate of cells is decreased by the addition of the test compound

[0081] Cells transformed with a nucleic acid molecule of the invention (histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:7) are useful as research tools. For example, one may obtain a cell (or a cell line, such as an immortalized cell culture or a primary cell culture) that does not express histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:8, insert a histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:7 nucleic acid molecule in the cell, and assess the level of expression and activity. Alternatively, histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:7 nucleic acid molecules may be over-expressed in a cell that expresses a histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:7 nucleic acid molecule. In another example, experimental groups of cells may be transformed with vectors containing different types of histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator SEQ ID NO:7 nucleic acid molecules to assess the levels of polypeptides and peptides produced, its functionality and the phenotype of the cells. The polypeptides and peptides are also useful for *in vitro* analysis of histidine SEQ ID NO:6 kinase, CSP SEQ ID NO:4 or response regulator SEQ ID NO:8 activity or structure. For example, the polypeptides and peptides produced can be used for microscopy or X-ray crystallography studies.

[0082] The histidine kinase SEQ ID NO:5 and SEQ ID NO:6, CSP SEQ ID NO:3 and SEQ ID NO:4 or response regulator SEQ ID NO:7 and SEQ ID NO:8 nucleic acid molecules and polypeptides are also useful in assays for the identification and development of compounds to inhibit and/or enhance polypeptide or peptide function directly. For example, they are useful in an assay for evaluating whether test compounds are capable of acting as antagonists for histidine kinase SEQ ID NO:6, CSP SEQ ID NO:4 or response regulator SEQ ID NO:8 by: (a) culturing cells containing a nucleic acid molecule which expresses histidine kinase SEQ ID NO:5, CSP SEQ ID NO:3 or response regulator peptides SEQ ID NO:7 (or fragments or variants thereof having histidine SEQ ID NO:6 kinase, CSP or response regulator activity) wherein the culturing is carried out in the presence of increasing concentrations of at least one test compound whose ability to inhibit histidine SEQ ID NO:6 kinase, CSP SEQ ID NO:4 or response regulator SEQ ID NO:8 is sought to be determined; and (b) monitoring in the cells the level of inhibition as a function of the concentration of the test compound, thereby indicating the ability of the test compound to inhibit histidine kinase SEQ ID NO:6, CSP SEQ ID NO:4 or response regulator SEQ ID NO:8 activity.

[0085] Biofilms are developed on polystyrene microtiter plates to provide a rapid and simple method for assaying biofilm formation, and hence activity of the peptide SEQ ID NO:4/receptor

[SEQ ID NO:8]/kinase **[SEQ ID NO:6]** system. Formation of biofilms is initiated by inoculating 20 μ l of cell suspension into each well containing 2 ml of biofilm medium (4X diluted Todd-Hewitt Yeast Extract supplemented with final concentration of 0.01% hog gastric mucin) for overnight incubation at 37°C under an anaerobic condition. After 20-h incubation, fluid medium is removed and added with 2 ml of pre-warmed, fresh THYE plus 5% horse serum. The cultures are incubated for 30 minutes and each well is supplemented with a final concentration of 200 ng/ml of synthetic competence stimulating peptide (SCSP) and varying concentrations of the inhibitor and the incubation is continued. After 30 minutes, plasmid DNA (1 mg/ml) or chromosomal DNA (10 mg/ml) is added to each well and the cultures are incubated for an additional 2 hr. Planktonic cells are then removed and the wells are washed once with PBS buffer. Biofilm cells are collected into 2 ml fresh medium by a gentle sonication or washing the wells using a pipette. The samples are centrifuged at 12,000 \times g for 5 min. Both biofilm and planktonic cells are resuspended into 200 μ l of fresh medium and are immediately spread on THYE agar plus appropriate antibiotics. Transformation frequency is determined after 48-h of incubation.

[0086] Homologues of the *Streptococcus pneumoniae* *comD* **[SEQ ID NO:5]**/**E** **[SEQ ID NO:7]** genes encoding a histidine kinase **[SEQ ID NO:6]**/ response regulator **[SEQ ID NO:8]** system were identified. This sequence was used to design primers to amplify the region from a number of *S. mutans* isolates. An open reading frame consisting of 138 nucleotides was located 148 nucleotides 5' proximal from the end of the *comD* homolog in the opposite orientation (Fig 1). This ORF was found to encode a peptide of 46-amino acid **[SEQ ID NO:2]** in length, the precursor of the 21-amino acid CSP **[SEQ ID NO:4]**.

[0087] The *comCDE* genes **[SEQ ID NO:23]** were amplified from the genomes of several *S. mutans* isolates by PCR using primers designed based on the genome database sequence and their nucleotide sequences determined. The deduced amino acid sequences are compared among the isolates by sequence alignment to confirm identity.

[0088] Genes are inactivated by integration of internal homologous fragments into the suicide vector pVA8912. Mutants defective in each of the individual genes (*comC* **[SEQ ID NO:1]**, *comD* **[SEQ ID NO:5]**, *comE* **[SEQ ID NO:7]**) are inactivated and their phenotypes are compared to the parent strain NG8 for their abilities to form biofilms, tolerate acidic pH (pH 2-4), and transport and incorporate DNA. The knockout mutants of *com D* **[SEQ ID NO:5]** and *E* **[SEQ ID NO:7]** were constructed by insertion-duplication mutagenesis, whereas the knockout *comC* **[SEQ ID NO:1]** mutant was created by allelic exchange via insertion of an erythromycin resistance determinant into the *comC* **[SEQ ID NO:1]** locus (Li *et al*, 2001). All mutant strains were therefore resistant to erythromycin. The wild-type strain was subcultured routinely on

Todd-Hewitt-Yeast Extract (THYE) agar plates (BBL®; Becton Dickinson, Cockeysville, MD), whereas the mutants were maintained on THYE agar plus 10 µg/ml of erythromycin. A minimal medium (DMM) was prepared to grow biofilms by a modification of the method described previously (Loo *et al*, 2000). The medium contained 58 mM K₂HPO₄, 15 mM KH₂PO₄, 10 mM (NH₄)₂SO₄, 35 mM NaCl, 2 mM MgSO₄·7H₂O, 0.2% (wt/vol) Casamino Acids and was supplemented with filter-sterilized vitamins, (0.04 mM nicotinic acid, 0.1 mM pyridoxine HCl, 0.01 mM pantothenic acid, 1 µM riboflavin, 0.3 µM thiamin HCl, and 0.05 µM D-biotin), amino acids (4 mM L-glutamic acid, 1 mM L-arginine HCl, 1.3 mM L-cysteine HCl, and 0.1 mM L-tryptophan) and 20 mM glucose.

[0090] To determine if the synthetic peptide [SEQ ID NO:16] could restore defective phenotypes of the *comC* [SEQ ID NO:2] mutants, a chemically synthesized 21-amino acid competence-stimulating peptide (CSP) [SEQ ID NO:4] (Li *et al*, 2001) was used in complementary experiments. The peptide was freshly dissolved in sterile distilled water to a concentration of 1 mg/ml. The CSP solution was then added to the cultures at a final concentration of 2 µg/ml 2 h after inoculation of bacterial cells.

[0096] Two methods modified based on the protocols described by Perry *et al* (Infect Immun, 41:722-727) and Lindler and Macrina (J Bacteriol, 166:658-665) were used to assay natural transformation of biofilm cells. Biofilms formed on polystyrene microtiter plates were added with 2 ml of pre-warmed, fresh THYE plus 5% horse serum (THYE-HS) immediately following removal of the BM medium, and the incubation continued at 37°C. After 2h incubation, a final concentration of 1 µg/ml plasmid DNA or 10 µg/ml of chromosomal DNA was added to each well. The cultures were incubated for an additional 2 h before collection of the cells for plating. To assay induction of competence by synthetic competence stimulating peptide (SCSP) [SEQ ID NO:16], the cultures were incubated for 30 min and a final concentration of 500 ng/ml of SCSP [SEQ ID NO:16] was added to each well. After a 30 min incubation, equal amounts of DNA was added to each well (1 µg/ml plasmid or 10 µg/ml of chromosomal DNA) and incubation continued for another 2 h. Fluid medium was then removed from individual wells and the wells were washed once with PBS buffer. Biofilm cells were collected into 2 ml PBS buffer by gentle sonication or by washing the wells using a pipette. The samples were centrifuged at 12,000 X g for 5 min. Both biofilm and planktonic cells were resuspended into 200 µl of fresh medium and were immediately spread on THYE agar plates plus appropriate antibiotics. For the biofilms developed in the chemostat, rods with biofilm cells were removed and placed into 2 ml of pre-warmed, fresh THYE-HS medium for 30 min incubation. Transformation was then

initiated by using the same methods as described above. The planktonic cells were also removed to compare the transformation frequency. After completion of the transformation procedures, both biofilm and planktonic cells were spread on THYE agar plus appropriate antibiotic. Transformation frequency was assessed after 48-h incubation. Transformation frequency was expressed as the number of transformants per μ g DNA per viable recipient at the time of DNA added.

In the Claims:

3. (Amended) The compound of claim 2, comprising a derivative of [SEQ ID NO:2] or [SEQ ID NO:4], a fragment of [SEQ ID NO:2] or [SEQ ID NO:4] or a derivative of a fragment of [SEQ ID NO:2] or [SEQ ID NO:4].
4. (Amended) The compound of claim 3, wherein amino acids are removed from the N-terminus and/or C-terminus of [SEQ ID NO:2] or [SEQ ID NO:4].
5. (Amended) A pharmaceutical composition comprising all or part of the peptide of ~~claims 1 to 4~~ claim 1 and a carrier.
6. (Amended) A method of medical treatment or prophylaxis of caries or endocarditis, comprising administering the compound of claim 1~~any of claims 1 to 4 or the pharmaceutical composition of claim 5~~.
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15. CANCELLED
16. CANCELLED
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18. CANCELLED
19. CANCELLED
20. CANCELLED
21. CANCELLED

24. (Amended) The polypeptide of claim 23 comprising all or part of an amino acid sequence in [SEQ ID NO:2] or [SEQ ID NO:4].

29. (Amended) An isolated nucleic acid molecule encoding the polypeptide of any of claims 24 22 to 28.

30. (Amended) An antibody directed against the polypeptide of any of claims 24 22 to 28.

32. (Amended) A vaccine composition comprising all or part of the peptide of any of claims 24 22 to 28 and a carrier.

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APPENDIX - 2

COMPLETE SET OF PENDING CLAIMS, AS AMENDED ON SEPTEMBER 17, 2001

1. A compound that competitively inhibits binding of CSP to *S. mutans* histidine kinase.
2. The compound of claim 1, comprising a peptide or an antibody.
3. (Amended) The compound of claim 2, comprising a derivative of [SEQ ID NO:2] or [SEQ ID NO:4], a fragment of [SEQ ID NO:2] or [SEQ ID NO:4] or a derivative of a fragment of [SEQ ID NO:2] or [SEQ ID NO:4].
4. (Amended) The compound of claim 3, wherein amino acids are removed from the N-terminus and/or C-terminus of [SEQ ID NO:2] or [SEQ ID NO:4].
5. (Amended) A pharmaceutical composition comprising all or part of the peptide of claim 1 and a carrier.
6. (Amended) A method of medical treatment or prophylaxis of caries or endocarditis, comprising administering the compound of claim 1.
22. An isolated CSP or a fragment thereof having *S. mutans* CSP activity.
23. The polypeptide of claim 22 comprising a *S. mutans* CSP.
24. (Amended) The polypeptide of claim 23 comprising all or part of an amino acid sequence in [SEQ ID NO:2] or [SEQ ID NO:4].
25. A polypeptide fragment of the peptide of claim 24, or a peptide mimetic of the CSP.
26. The polypeptide of claim 24 which is recombinantly produced.
27. A polypeptide comprising a sequence having greater than 30%, 50% or 60% sequence identity to the polypeptide of claim 24.
28. The polypeptide of claim 24, isolated from *S. mutans*.
29. (Amended) An isolated nucleic acid molecule encoding the polypeptide of any of claims 22 to 28.
30. (Amended) An antibody directed against the polypeptide of any of claims 22 to 28.
31. The antibody of claim 30, comprising a monoclonal antibody or a polyclonal antibody.
32. (Amended) A vaccine composition comprising all or part of the peptide of any of claims 22 to 28 and a carrier.
33. The vaccine composition of claim 32, wherein the peptide is coupled to a compound comprising all or part of KLH, ovalbumin, or thyroglobulin.

APPENDIX - 3

DETAILS OF DRAWING REVISIONS SUBMITTED ON SEPTEMBER 17, 2001

Express Mail No. EL818623952US

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Figure 1

Schematic layout of the arrangement of the genetic locus encoding the signal peptide-precursor, the histidine kinase and the response regulator. Note that this arrangement is different from other loci in related streptococci for the following reasons: a) The *comC* gene is transcribed from its own promoter alone, unlike the genes thus far described in other streptococci that are arranged in an operon-like cluster with the *comC/DE* genes being transcribed from a single promoter. b) The *comC* gene is separated from the *comD* gene by 148 nucleotides.

Streptococcus mutans
ComCDE Operon



Figure 2

Sequences of the open reading frames encoding the signal peptide precursor (ComC), the histidine kinase (ComD), and the response regulator (ComE).

A.

→ *S. mutans* comC gene

Encodes a precursor to a signal peptide

[ATGAAAAAAACACTATCATTAAAAATGACTTTAAAGAAATTAAAGACTGATGAATTAG
AGATTATCATTGGCGGA (AGCGGAAGCCTATCAACATTTCGGCTGTTAACAGAAG
TTTACACAAGCTTGGGAAAA)] TAA

B.

→ *S. mutans* CSP encoding sequence

Competence Signal Peptide

AGCGGAAGCCTATCAACATTTCGGCTGTTAACAGAAGTTTACACAAGCTTGGG
AAAA [SEQ ID NO:1]

C.

→ *S. mutans* comD gene

Encodes a protein that functions as a histidine kinase receptor

[ATGAATGAAGCCTTAATGATACTTTCAAATGGTTATTAACCTATCTAACCGTTCTAT
TTCTCTGTTCTATTTCTAAGGTAAGTAATGTCACTTATCGAAAAAGGAATTAACT
CTTTTTCTGATAAGCAATTTCGATAATGATTGCTGTTACGATGGTGAACGTAAACCT
GTTTATCCTGCAGAGCCTTTATTTATAGCTTATCAATTATCTTAATAGACAGA
ATAGTCTTCTCTAAATATATTATGGTCTGCTGCCTGTTGCCAGTTCTGACTTGT
AGGCGGGCAATCATATTCTTATCTGGATGAACTCAAGGAATTGTAATGGCAGTAG
CATTATAACCACCTATATGATCGAGTTGCAGGAATAGCGCTAAGTTACCTCTTC
GTGTGTTCAATGTTGATATTGGTCGACTAAAGATAGTTGACCAAGATGAAGGTCAAA
AAACGCTTGATTCCAATGAATATTACTATGCTTCTATACTACCTTTAAACAGGTATT
GTATGTTATAGAGAGTTATAATGTGATACCGACTTTAAATTTCGTAATTGTCGTTA
TTGTCTATCTTATTATTTGATTCTGATCTCATTTTAAGCCAATATACCAAACAA
AAGGTTCAAAATGAGATAATGGCACAAAGGAAGCTCAGATTGAAATATCACCCAGTA
TAGTCAGCAAATAGAATCTCTTACAAGGATATTGAGTTCCGCCATGATTATCTGA
ATATTTAACTAGCCTCAGATTAGGCATGAAAATAAGATTAGCTAGTATTGAAAAG
ATTTACCATCAAATCTAGAAAAACAGGACATCAATTGCGAGGATACCGTTATAATAT
CGGCCATCTAGCTAATATTCAAAACGATGCTGTCAGGGTATCTGTCAGCAAAATCT
TAGAAGCTCAGAATAAAAGATTGCTGTCATGAGAAGTCTCAAGTAAATACAACCTG
CCTGAGATGGAGTTGCTGATTCTTACCATACCTTCTATCTTGTGATAATGCCAT
TGAGGCTGCTTCGAATCATTAATCTGAAATTCACTGTTAGCCTTTTAAGAAAAATG
GCAGTATAGTCTTATCATTGAGATTCCACCAAGAAAACAATAGATGTGAGTAAA
ATTTTAAAGAAAATTCACAAAGGCTCCATCGCGGTATTGGTTAGCAAAGGT
GAATCATATTCTGAAACATTCCAAACCAAGTTACAAACAAAGCAATCATCATCATT
TATTCAAGCAACTCCTAATAATAAA] TAG

D.

→ *S. mutans* comE gene

Encodes a response regulator that activates transcription of a number of genes

Figure 2 (cont'd)

[ATGATTCTATTTGATTGAAAGATGATTTTACAACAAGGACGTCTGAAACCA
CCATTGCAGCTATCATGAAAGAAAAAATTGGCTTATAAAGAATTGACTATTTGGA
AAACCACAACAACCTATTGACGCTATCCCTGAAAAGGGCAATCACCAGATTTCTTTT
GGATATTGAAATCAAAAAGAGGAAAAGAAAGGACTGGAAGTAGCCAATCAGATTAGAC
AGCATAATCCTAGTGCAGTTATTGCTTTGTCACGACACATTCTGAGTTATGCCCTC
ACTTTTCAGTATCAGGTATCTGCTTGGATTATTGATAAAATCTTGAATCCTGAGGA
GTTCTCCCACCGCATTGAATCAGCGCTGTATTATGCTATGGAAAACAGCCAGAAGAATG
GTCAATCAGAGGAACCTTTAATTCCATTCACTGAAACTCAGTTCAGGTCCCTTTT
GCTGAGATTCTGTATTTGAAACATCTCAACAGCCCATAAGCTCTGCCTTATACTTA
TGATGAACGGATTGAATTCTACGGCAGTATGACTGACATTGTTAAAATGGATAAGAGAC
TTTTCAAGTGCCATCGCTTTTATTGTCATCCTGCCAATATTACCGTATTGATCGG
AAAAAACGCTTGGCTATTTGAAATAATAAGTCTTGTCTTATTCAACGAACTAAGTT
AACAAAACGTGAGAGCTGTGATTGCTGATCAAAGGAGAGCAAA] TGA

Figure 3

The amino acid sequences of the signal peptide precursor (ComC), the histidine kinase (ComD), and the response regulator (ComE).

4.

➤ S. mutans ComC protein (CSP precursor)

MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK

5.

➤ S. mutans ComD protein (Histidine Kinase)

MNEALMILSNGLLTYLTVLFLFLSKVSNVTLKKELTLFSISNFLIMIAVTMVNVNL
FYPAEPLYFIALSIYLNQRQNSLSLNIFYGLLPVASSDLFRRAIIFFILDGTQGIVMGSS
IITTYMIEFAGIALSYLFLSVFNVDIGRLKDSLTKMKVKRLLIPMNITMLIYLLIQVL
YVIESYNVIPTLKFRKFVVIVYLFLILISFLSQYTKQKVQNEIMAQKEAQRNITQY
SQQIESLYKDIRSFRHDYLNILTSRLGIENKDLASIEKIYHQILEKTGHQQLQDTRYNI
GHLANIQNDAVKGILSAKILEAQNKIAVNVEVSSKIQLPEMELLDFITILSILCDNAI
EAAFESLNPEIQLAFFKKNGSIVFIIQNSTKEKQIDVSKIFKENYSTKGSNRGIGLAKV
NHILEHYPKTSQTSNHHHLFKQLLIIK

6.

➤ S. mutans ComE protein (Response Regulator)

MISIFVLEDDFLQQGRLETTIAAIMKEKNWSYKELTIFGKPQQLIDAYPEKGNHQIFFL
DIEIKKEEKKGLEVANQIRQHNPSAVIVFVTTSEFMPLTQYQVSALDFIDKSLNPEE
FSHRIESALYYAMENSQKNGQSEELFIFHSSETQFQVPPFAEILYFETSSTAHKLCLYTY
DERIEFYGSMTDIVKMDKRLFQCHRSFIVNPANITRIDRKRLAYFRNNKSCLISRTKL
TKLRAVIADQRRAK

Figure 4

The deduced amino acid sequence of the signal peptide precursor in various strains and its predicted cleavage site. The original peptide is expressed as a 46 amino acid peptide that is cleaved after the glycine-glycine residues to generate an active signal peptide.

BM71 CSP	1 MKKTPSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
GB14 CSP	1 MKKTLSLKNDFKSIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
E7 CSP	1 MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
JH1005 CSP	1 MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
LT11 CSP	1 MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
NG8 CSP	1 MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46
UAB159 CSP	1 MKKTLSLKNDFKEIKTDELEIIIGGSGSLSTFFRLFNRSFTQALGK	46

consensus: 1 MKKTLSLKNDFKEIKTDELEIIIGG SGSLSTFFRLFNRSFTQALGK 46
predicted cleavage site:

Figure 5

The synthetic signal peptide that is effective at inducing competence, biofilm formation and acid tolerance in *Streptococcus mutans*.

SGSLSTFFRLFNRSFTQALGK [SEQ ID NO:2]

Figure 6

The natural activity of the signal/receptor system functioning *in vitro* in model biofilms
as determined by the ability of various strains of *S. mutans* to accept donor plasmid DNA
conferring erythromycin resistance.

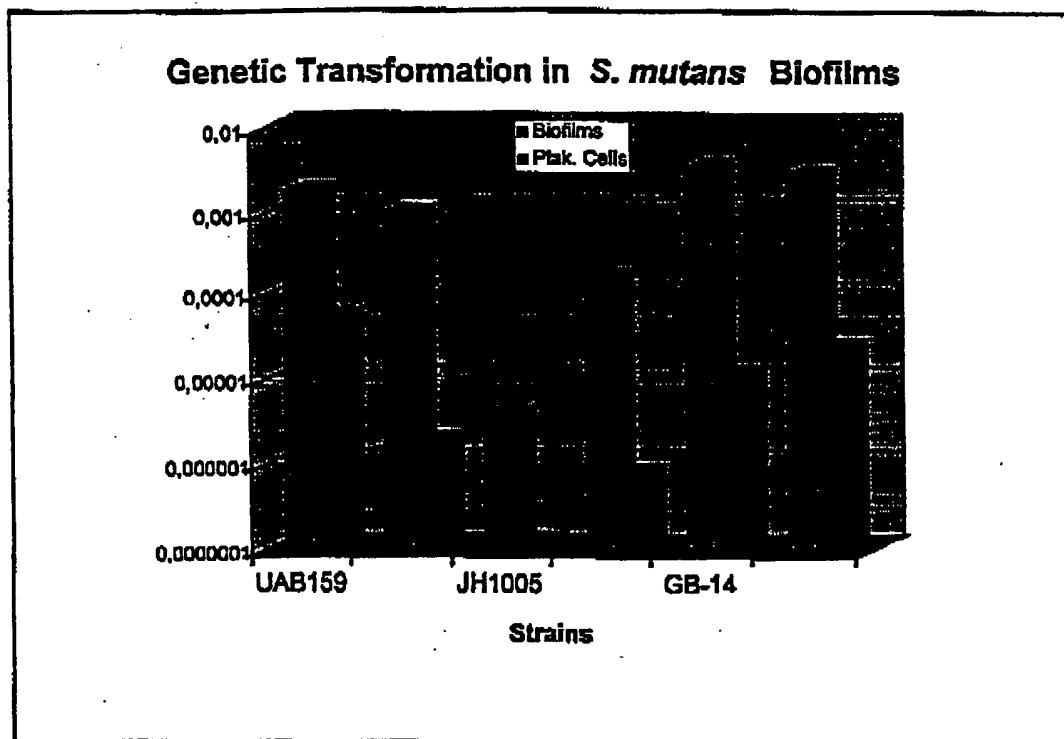


Figure 7
Induction of genetic transformation in *Streptococcus mutans*
by synthetic competence stimulating peptide (SCSP)¹

Strain	Peptide added Number of Transformants/Recipients	No peptide Number of Transformants/Recipients
UAB15	4.65×10^{-1}	1.78×10^{-6}
JH1005 ²	6.98×10^{-2}	0

¹The final concentration of SCSP used was 500 ng/ml.

The strain contains a nonsense mutation in the *comC* gene encoding the CSP.

Figure 8

~~List of the primers used to amplify the genes or internal regions of the target genes by polymerase chain reaction (PCR) for subsequent sequencing or inactivation.~~

ComC region

ComC Primer Pair: F5-B5

[F5] 23406-23424 5'- AGTTTTTGTCTGGCTGCG -3'

19 nt forward primer

pct G+C: 47.4 Tm: 50.5

[B5] 24056-24037 5'- TCCACTAAAGGCTCCAATCG -3'

20 nt backward primer

pct G+C: 50.0 Tm: 51.9

651 nt product for F5-B5 pair (23406-24056)

Optimal annealing temp: 50.3

pct G+C: 30.9 Tm: 71.5

ComD region

ComD Primer Pair: F1-B1

[F1] 392-415 5'- CGCTAAGTTACCTCTTCAGTG -3'

24 nt forward primer

pct G+C: 45.8 Tm: 51.6

[B1] 683-663 5'- GCTTCCTTGTGCCATTATC -3'

21 nt backward primer

pct G+C: 42.9 Tm: 50.8

292 nt product for F1-B1 pair (392-683)

Optimal annealing temp: 49.5

pct G+C: 30.8 Tm: 70.2

ComE region

ComE Primer Pair: F1-B1

[F1] 145-165 5'- CCTGAAAAGGGCAATCACCAG -3'

21 nt forward primer

pct G+C: 52.4 Tm: 55.9

[B1] 606-585 5'- GCGATGGCACTGAAAAAGTCTC -3'

22 nt backward primer

pct G+C: 50.0 Tm: 55.4

462 nt product for F1-B1 pair (145-606)

Optimal annealing temp: 53.6

pct G+C: 38.3 Tm: 74.1

Figure 9

ComCDE local region. The ComC (first highlighted region; nucleotides 101 to 241), ComD (second highlighted region; nucleotides 383 to 1708) and ComE (third highlighted region; nucleotides 1705 to 2457) proteins are highlighted.

Sequence Range: 1 to 2557

10 20 30 40 50
 ACATTATGTCCTAAGGAAAATATTACCTTTCAAGAAAATCCATGATT
 TGTAATACACAGGATTCTTTATAATGAAAAAGTTCTTTAGGTACTAA
 <K K L F I W S K

60 70 80 90 100
 TTTCATAAAAATAGTATACTAATTATAATCAAAAAAGGAGATATAAA
 AAAAGTATTTTATCATATGATTAATTAGTTTTCTCTATATTT
 <K M F F L I S I I I L F L L Y L

110 120 130 140 150
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 TACTTTTTGTGATAGTAATTCTACTGAAATTTCTTTAATTCTGACT
 <K M F F L I S I I I L F L L Y L
 ORF RF [2]

160 170 180 190 200
 TGAATTAGAGATTATCATCGCGGAAGCGGAAGCCTATCAGATTTCC
 ACTTAATCTCTAATAGTAACCGCCCTCGCCCTCGGATAGTTGTAAAAAGG
 <S N S I I M
 ORF RF [2]

210 220 230 240 250
 GGCTGTTAACAGAAAGTTTACACAAAGCTTGGAAAAATAAGATAGGCTA
 CCGACAAATTGTCCTAAATGTCGAAACCCCTTTATCTATCCGAT
 <S N S I I M
 ORF RF [2]

260 270 280 290 300
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 TGTAACCTTATTITGTCCGACCTAATAATAAGGTCCGAAAAATTACA

310 320 330 340 350
 AAAATAAAATACAGGGTAAATAATCAAGTGTGCTGTCGTGGATGAGAA
 TTTTATTTATGTCCCATTATTAGTTACACAGCACGACACTACTCTT

360 370 380 390 400
 GATAAAACTATCTCTTAGAGAAATAGGCCTCTATTTTATTATTAGGAG
 CTATTITGATAGAGAAATCTCTTACCGGAGGAGATAAAATAATAATCCTC
 <S N S I I M
 ORF RF [1]

410 420 430 440 450

Figure 9 (cont'd -1)

AACGAACCTAATTACTACTAACGAACAAACATTGACCAAAACCTA
 < ORF RF [4] C >

460 470 480 490 500
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 < H E L I H N V K A L G I G R N S
 < ORF RF [4] C >

510 520 530 540 550
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 GGAAATCACCTTATCAAAGAAATTCTAAATGAGTGTAGATAAACAAA
 < G W K N T S V N A E K P R T V T S V D E S N B O K
 < ORF RF [4] C >

560 570 580 590 600
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 AAGAAACCACCTTAAGACTTACTATTCTGATATGACGGTAAAAGAATT
 < E K E T S N O D T P V V T S G N T K K G T E
 < ORF RF [4] C >

610 620 630 640 650
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 TTTCCGATTGACTTAAAGTCTAAATTACTAAGCTTCGTCGGAGTAC
 < M >

< ORF RF [4] C >

660 670 680 690 700
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 < A L S H K I E S M V M K S S N S I >
 < ORF RF [3] >

< ORF RF [4] C >

710 720 730 740 750
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 GAGTCGGTCAACATAAAATGAACCTCTGAAAGATGAACTGTCGTTAGAAAA
 < S G S C I L L E T S T L T A I F >
 < ORF RF [3] >

< ORF RF [4] C >

760 770 780 790 800
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 < L F >

< ORF RF [4] C >

810 820 830 840 850
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Figure 9 (cont'd-2)

ATGTCCTGTTTCTAAGATTTGATGGTAAATCTTTCAATACTAGCTA
 TACAGGACAAAAAGATTCTAACTTACCAATTAGAAAAGTTATGATCGAT
 ← ORF RF [4] C →

860 870 880 890 900
 ATGTCCCTGTTTCTAAGATTTGATGGTAAATCTTTCAATACTAGCTA
 TACAGGACAAAAAGATTCTAACTTACCAATTAGAAAAGTTATGATCGAT
 ← ORF RF [4] C →

910 920 930 940 950
 AATCTTTATTTCAATGCCTAATCTGAGGCTAGTTAAAATATTCAAGATAA
 TTACAAAATAAAAGTTACCGATTAGACTCCGATCAATTATAAGTCTATT
 ← ORF RF [4] C →

960 970 980 990 1000
 TCATGGCGAAACTCGAATATCCTTGTAAAGAGAATTCTATTGCTGACT
 AGTACCGCCTTGAAGCTTATAGGAACATTCTCTAAGATAAACGGACTGA
 M A E T S N I L V K R F Y L L T >

← D H R E S T R I D K Y N S E D H O O S
 ORF RF [4] C →

1010 1020 1030 1040 1050
 ATACTGGGTGATATTCGAATCTGAGCTTCTTTGTGCCATTATCTCAT
 TATGACCCACTATAAGCTTAGACTCGAAGGAAAACACGGTAATAGAGTA
 I L G D I S N L S F L L C H Y L I >

← E G T E I N R E I O Q A H E K O T H M T E N
 ORF RF [4] C →

1060 1070 1080 1090 1100
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 AAACCTGGAAAACAAACCATATAACCGAATTCTACTCTAGCTTAGTT
 L N L L F G I L A >

← Q V E K O F T Y N O S E E F S C I A E E L
 ORF RF [4] C →

1110 1120 1130 1140 1150
 AATAAAATAAGATAGACAATAACGACAAATTACGAATTAAAGTCGG
 TTATTTTATTCTATCTGTATTGCTTAAATGCTTTAAAATTTCAAGCC
 ← F D L D Y V D V V E E K R F K L T >
 ORF RF [4] C →

1160 1170 1180 1190 1200
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 ← G V N Y S E I V Y L V O I L I V Y Y
 ORF RF [4] C →

1210 1220 1230 1240 1250
 ATAGAAGCATAGTAATAATCATTGGAAATCAAGCGTTTTGACCTTCATC
 TATCCTCGTATCATTATAACTAACCTTAGTTGCGAAAAACTGGAAAGTAG
 ← G V L M T I N M E E E P R G K V K C M
 ORF RF [4] C →

Figure 9 (contd-3)

1260 1270 1280 1290 1300
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 AACAGTTGATAGAAATTCTAGCTGGTTATAGTTGAACTTGTCGACTC
 < K T Y I T S D K P R G C E D P V V N S P V V S L
 ORF RF [4] C

1310 1320 1330 1340 1350
 AAAAGGGTAACTTAGCGCTATTCTGCAAACCTCGATCATATAGGTGGTTA
 TTCTCCATTGAATCGCGATAAGGACGTTGAGCTAGTATATCCACCAAT
 < P T I L V E N S I A A T T G G A A P E E V E E V E E S T H
 ORF RF [4] C

1360 1370 1380 1390 1400
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 ATTACGATGACGGGTAATGTTAAAGGAACTCAAGGTAGGTTCTATTTCTTA
 < G T S S G M V I G Q T C G C D S L A T E P F
 ORF RF [4] C
 < L E K L E M W S L S Y

1410 1420 1430 1440 1450
 ATGATTGCCGCCCTAAACAAAGTCAGAACTGGCAACAGGCAGCAGACCAAA
 TACTAACGGGGGATTTCAGTCAGTCAGTCAGCTTGACCGTTGTCGTCTGGTAT
 < K T T A A P R C R E V L D D S S S A A T V V E E V E E G V X
 ORF RF [4] C
 < S Q G G L C T L V P L L C C V M

1460 1470 1480 1490 1500
 AAATATATTTAGAGAAAAGACTATTCTGTCTTAAAGATAATTGATAAAG
 TTTATATAAATCTCTTCTGATAAGACAGATAAATCTATTAACTATTTC
 < P T I N D S K L S M V O R N N E E V E E S V D W S
 ORF RF [4] C

1510 1520 1530 1540 1550
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 GATATTTTATTTCTCGAGACGTCTTAAATGCAAGTGGTAG
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 ORF RF [4] C

1560 1570 1580 1590 1600
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 CATTGTCGTTAGTAATGCTTTAAAGAATAGCTTTCTCAATTAAAG
 < T V Y A T I M L L K N S V S V S V L T E E E E E E
 ORF RF [4] C

1610 1620 1630 1640 1650
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 GAAAAAGCTATTTCAGTGAATGAAATGGAATCTTTATCTTTGTTCTCTT
 < K T K L V T V N S V V K S V S V L T E E E E E E
 ORF RF [4] C

1660 1670 1680 1690 1700
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 < K T V T E V Y T L E G N S V S V L T E E E E E E
 ORF RF [4] C

Express Mail No. EL818623952US
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Atty Dkt No.: 1889-00401
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Figure 9 (contd-4)

1710 1720 1730 1740 1750
TCATTCATTTGCTCTCTTGTACAGCAATCACAGCTCTCAGTTTTGTT
AGTAAGTAAACGAGAGGAAACTAGTCGTTAGTGTGAGAGTCAAAACAA
M
ORF RF [5] C

1760 1770 1780 1790 1800
AACTTAGTCGTGAAATAAGACAAGACTTATTATTCGAAAATAGGCCAA
TTGAATCAAGCACTTTATTCTGTTCTGAATAATAAGCTTATCCGGTT
C
ORF RF [5] C

1810 1820 1830 1840 1850
GGGTTTTCCGATCAATACGGTAATATTGGCAGGATTGACAATAAAAG
CGCAAAAAAGGCTAGTTATGCCATTATAACCGCTCTAAGTGTATTTC
C
ORF RF [5] C

1860 1870 1880 1890 1900
AGCGATGGCACTGAAAAAGTCTCTTATCCATTAAACATGTCAGTCATA
TCGCTACCGTGACTTTTCAGAGAATAGGTAAAATTGTTACAGTCAGTAT
M A L K K S L I H F N N V S H
ORF RF [1] >
ORF RF [5] C
<V
<

1910 1920 1930 1940 1950
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GACGGCATCTTAAGTTAGCAAGTGTATTCAATTTCGGCTCTCGAATAC
T A V E F N P F I I S I K A E L M
ORF RF [1] >
ORF RF [5] C
<A T S N L G N M M L I F A S S I
<ORF RF [6] C

1960 1970 1980 1990 2000
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CCGACAACCTCTACAAAGTTATGCTTAGAGTCGTTCCCTGGACTT
G C
>
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<ORF RF [5] C
<P Q Q L H K L I C F R L L L S R F
<ORF RF [6] C

2010 2020 2030 2040 2050
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S O T T E S S H F I F A L P B E R S A Q G I N
<ORF RF [5] C
<S L K L H I S F L F N R Q N V M R

Figure 9 (cont'd-5)

ORF RF[6] C

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AAGACCGACAAAGGTATCGTATTATGTCGCGACTAAGTTACGCCACCC
S> < R D K M A Y V I A S S N D E P R K S

ORF RF[5] C

< R A T K W L M
< ORF RF[6] C

2110 2120 2130 2140 2150

GAACCTCCTCAGGATTCAAAGATTATCAATAAAATCCAAGCAGATACT
CTTGAGGAGTCCTAAGTTCTAAATAGTTATTAGGTTTCGTCTATGGA
S> < R D K M A Y V I A S S N D E P R K S

ORF RF[5] C

2160 2170 2180 2190 2200

GATACTGAAAAGTGAGGGCATAAAACTCAGAATGTGTCGTGACAAAGACA
CTATGACTTTCACTCCCCTATTTGAGTCCTACACAGCACTGTTCTGT
M C R D K D >

< R D K M A Y V I A S S N D E P R K S
< ORF RF[5] C

2210 2220 2230 2240 2250

ATAACTGCACTAGGATTATGCTGTCATAATCTGATTGGCTACTTCCAGTCC
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N N C T R I M L S N L I G Y F Q S >

< R D K M A Y V I A S S N D E P R K S
< ORF RF[5] C

2260 2270 2280 2290 2300

TTTCTTTCTCTTTTGTGATTCAATATCCAAGAAATCTGGTGA
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F L F L F F D F N I Q K E N L V I >

< R D K M A Y V I A S S N D E P R K S
< ORF RF[5] C

2310 2320 2330 2340 2350

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ACGGGAAAAGTCCCTATCGCAGTTATTCAACAAACACCAAAAGGTTTAT
A L F R D S V N K L L W F S X N >

< R D K M A Y V I A S S N D E P R K S
< ORF RF[5] C

2360 2370 2380 2390 2400

GTCAATTCTTATAAGACCAATTCTTCTTGTGATAGCTGCAATGGT
CAGTTAAGAAATATTCTGGTAAAGAAAGTACTATCGACGTTACCA
S Q F F I R P I F F F H D S C N G >

< R D K M A Y V I A S S N D E P R K S
< ORF RF[5] C

Figure 9 (cont'd-6)

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G P K T S L I >
V S R R P C C K K S S S N T K I >
S T E M D C R G Q Q S I P L E D M D D E L V A E P V S
< ORF RF [5] C >
2460 2470 2480 2490 2500
AAATCATTATTCTCCCTTAATCTCTATTAGTTAGCTGATTAACACT
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S I I I S P L I F Y L G >
< S T E M D C R G Q Q S I P L E D M D D E L V A E P V S
2510 2520 2530 2540 2550
ATACACAGAAAAGGTATAAACGATATCACTCAATAAAATCTACTAACCT
TATGTGCTTTCCATATTGCTATAGTGTAGTATTAAATGATTGAA
AATAACC
TTATTGG

Figure 10

The comX nucleotide sequence, amino acid sequence, and its local region with 100bp included both upstream and downstream (promoter is upstream).

(A)

➤ S. mutans comX gene

ATGGAAGAAGATTGAAATTGTTTAATAAGGTTAACGCCATTGTATGGAAATTAAAG
CCGTTATTACTTATTAAATGTGGACTCGTGAAGATTGGCAACAAGAGGGAAATGTTGA
TTTGCAACCAATTATTAAGGGAACATCCAGAATTAGAAGAGGGATGATAACAAATTGTAT
ATCTATTAAAGACACGTTCTAATTACATTAAAGATGTTGCGTCAGCAAGAAAG
TCAGAAACGTCGTTAAATAGAATGTCTTATGAAGAAGTCGGTGAGATTGAACACTGTT
TGTCAAGTGGCGGTATGCAATTGGATGAATATATTAACTTGTGATAGTTGCTTGCA
TATAAACAAAGGCTGAGTACTGAAAGCAAGAGCTGTTGAGGCCCTGGTACCGAGGAGA
GCACTTTGGGAAGGCAAAGTATGCTGAAAGATTACGTAAAAAAATTAAAGTGATTAA
AGGAAAAA

(B)

➤ S. mutans ComX protein

MEEDFEIVFNKVVKPIVWKLSRYYFIKMWTREDWQQEGMLILHQLLREHPELEEDDKLY
IYFKTRFSNYIKDVLRQQESQKRRFNRMSYEEVGEIEHCLSSGGMQLDEYILFRDSLLA
YKQGLSTEKQELFERLVAGEHFLGRQSMLKDLRKKLSDFKEK

(C)

➤ S. mutans comX gene local region

GTAAATAAAACAGCCAGTTAACGATGGACATTATGTCCTGTTCTAAAGCTTTTCG
TTTATAATAATTATTATAAAAGGAGGTATCGTAATAGATGGAAGAAGATTGAA
ATTGTTTAATAAGGTTAACGCAATTGTATGGAAATTAAAGCCGTTATTACTTTATTAA
AATGTGGACTCGTGAAGATTGGCAACAAGAGGGAAATGTTGATTTGACCAATTATTAA
GGGAACATCCAGAATTAGAAGAGGGATGATACAAAATTGTATATCTATTAAAGACACGT
TTTCTAATTACATTAAAGATGTTGCGTCAGCAAGAAAGTCAGAAACGTCGTTAA
TAGAATGTCTTATGAAGAAGTCGGTGAGATTGAACACTGTTGTCAGTGGCGGTATGC
AATTGGATGAATATATTAACTTGTGATAGTTGCTTGATATAAAACAAGGTCTGAGT
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AAAGTATGCTGAAAGATTACGTAAAAAAATTAAAGTGAATTAAAGGAAAAATAGTTAAAAA
GGGAAAGAATGGAACATGTGATTGTACCAATTGTTGAAATTAAAGAAAAGTTA
TTATAAAATTATTGGTTAACATGCCATATTAA

Figure 11.

~~The *comA* and *comB* nucleotide and amino acid sequences. *ComA* and *ComB* are the components of the CSP exporter.~~

(A.)

S. mutans *comA* gene

```
ATGAAACAAGTTATTTATGTTGTTTAATCGTCATAGCCGTTAACATTCTCTAGAGAT
TATCAAAAGAGTAACAAAAGGGGAGGGACAGTTCGTCATCTAACCTTACCAAGATG
GGCAGTCTAAGTTGTTGCGCAGACATTATAAGCTAGTACCTCAGATTGATACCGA
GACTGTGGGCCGGCAGTGTGGCATCTGTTGCAAAGCATTACGGATCTAATTACTCTAT
CGCTTATCTGCGGAACTCTCAAAGACTAACAAAGCAGGGAAACAACAGCTCTGGCATTG
TTGAAGCTGCTAAAAAGTTAGGCTTGAAACACGGCTCTATCAAGGGGATATGACGCTT
TTTGATTATAATGATTGACCTATCCTTATCGTCATGTGATTAAAGGAAAACGCT
GCAGCATTATTATGTCGTCTATGGCAGCCAGAATAATCAGCTGATTATTGGAGATCTG
ATCCTTCAGTTAAGGTGACTAGGATGAGTAAGGAACGGCTTCAATCAGAGTGGACAGGC
CTTGCAATTTCCTAGCTCTCAGCTTAACCTATAAGCCTCATAAAGGTGAAAAAAATGG
TTTGTCTAATTCTCCCGTTGATCTTAAGCAGAAAGCTTGATGACTTATATTATCA
TAGCTAGCTTGATTGTGACGCTCATTGATATTGTCGGATCATACTATCTCCAAGGAATA
TTGGACGAGTACATTCCGTCTGATCAGCTGATTCAACTTTAGGAATGATTACGATTGGTCT
GATAATAACCTATATTATCCAGCAGGTCTGGCTTTGCAAAAGAATACCTCTGGCCG
TACTCAGTTGCGTTAGTCATTGATGTTATCCTGTCTTATATCAAACATATTTTACG
CTTCCTATGTCTTCTTGCAGAACAGGGAACAGGAGAAATCAGCTCTGTTTACAGA
TGCCAATCAGATTATTGATGCTGTAGCGTCAACCCTTTCAATCTTTAGATATGA
CTATGGTAATTGGTTGGGGTTTGTGGCGAAAACAATAACCTTTCTTCTA
ACCTTGCTCTCCATTCCGATTATGCCATCATTATTTGCTTCTTGAAACCTTGA
GAAAATGAATCAGAACGATGGAAAGCAATGCTGTGGTAAGTCTTCTATCATTGAAG
ATATCAATGGGATGAAACCTAAATCACTCACAAGTGAATCCGCTCGTTATCAAAC
ATTGATAGTGAATTGTTGATTATTGGAGAAAACCTTAAGCTACACAAGTATAGTC
CATTCAAACCGCATTAAAAGCGGTGCTAACGTTATCCTCAATGTTGTCATTCTGGT
ATGGCTCTCGTCTAGTTATGGATAATAAAATCTCAGTTGGTCAGCTTATCACCTTAA
GCTTGTGCTCTTATTCCTCAAATCCAATTGAAAATATTATCAAATCTGCAATCCAAC
GCAGTCAGCTCGCGTTGCCAATCACAGCTTAATGAGGTCTATCTGCGAACTGAAT
TTGAAAAAGACGGCATTATCAGAAAATAGCTTTAGATGGTGAATTTCGTTGAA
AATCTTCTTATAAATATGGATTGGCGAGATAACCTTATCAGATAATTATCAAT
CAAAAAGGCTCCAAGGTCAAGTCTAGTTGGAGGCCAGTGGTCTGGTAAAACAACCTGG
CTAAACTGATTGTCATTCTACGAGCCTAACAAAGGGATTGTTGAAATCAATGGCAAT
GATTAAAAGTTATTGATAAGACAGCTTGGCGGCGATATTAGCTATTGCGCAACA
GGCCTATGTTTAGTGGCTCTATTATGGATAATCTGTTTAGGAGCTAAAGAAGGAA
CGAGTCAGGAAGACATTATCGTGTGAAATTGCTGAAATCCGCTCGGACATTGAA
CAAATGCCCTAGGGCTATCAGACAGAGTTATCAGATGGTCCGGTATTCTGGCGGTCA
AAAACAGCGGATTGCTTCTAGGGCTTATTAACACAGGCACCGGTTTGATTCTGG
ATGAAGCCACCAGCAGTCITGATAATTGACAGAAAAGAAAATTATCAGCAATCTCTTA
CAGATGACGGAGAAAACAATAATTGGTGTGCCACCGCTTAAGCATTACAGCGTAC
TGACGAAGTCATTGTCATGGATCAGGGAAAATTGTTGAACAAGGCACTCATAAGGAAC
TTTAGCTAAGCAAGGTTCTATTATAACCTGTTAAT
```

(B.)

S. mutans *ComA* protein

Figure 11 (cont'd)

(B.)

MKQVITYVVLIVIAVNILLEIIKRVTKRGGTVSSSNPLPDGQSKLFWRRHYKLVPQIDTR
 DCGPAVLASVAKHYGSNYSIAYLRELSKTNKGTTALGIVEAAKKLGFETRSIKADMTL
 FDYNDLTYPFIVHVIKGKRLQHYYVYGSQNNQLIIGDPDPSPVKVTRMSKERFQSEWTG
 LAIFLAPQPNYKPHGEKNGLSNFFPLIFKQKALMTYIIIASLIVTLIDIVGSYLYQGI
 LDEYIPDQLISTLGMITIGLITYIIQVMAFAKEYLLAVLSRLVIDVILSYIKHIFT
 LPMSFFATRRTGEITSRFTDANQIIDAVASTIFSIFLDMTMVLVGGVLLAQNNNNLFFL
 TLLSIPYAIIFIAPFLKPFEMNHVEMESNAVSSSIIEDINGMETIKSLTSESARYQN
 IDSEFDVYLEKNFKLHKYSATQALKSGAKLILNVILWYGSRLVMDNKISVGQLITFN
 ALLSYFSNPIENIINLQSKLQSARVANTRLINEVYLVESEFEKDGDLSENSFLGDISFE
 NLSYKYGFGRDLS DINLSIKGSKVSLVGASGSGKTTLAKLIVNFYEPNKGIVRINGN
 DLKVIDKTALRRHISYLPQQAYVFSGSIMDNLVLGAKEGTSQEDIIRACEIAEIRSDIE
 QMPQGYQTELSDGAGISGGQKQRIALARALLQAPVLLDEATSSLILTEKKIISNLL
 QMTEKTIIFVAHRLSISQRTDEVIVMDQGKIVEQGTHKELLAKQGFYYNLFN

(C.)

S. mutans *comB* gene

ATGGATCCTAAATTTCACAAAGTCAGAAATTATAGGAGACGCTATCATAATTTCGC
 GACACTATTAATTGTTCTTGGCTGCTGATTATCTTCTGGTCATATTCTTTGTT
 TTGCTAAAAAGAAATTACAGTGATTCTACTGGTGAAGTTGCACCAACAAAGGTTGTA
 GATGTTATCCAATCTTACAGTGACAGTTCAATCATAAAAATAATTAGATAATAATGC
 AGCTGTTGAGAAGGGAGACGTTTAATTGAATATTCAAGAAAATGCCAGTCAAACCGTC
 AGACTGAACAAAAGAAATTATAAAAGAAACAAAAGAGAAGAGAAGAGAAGGAAAAGAAA
 AAACACCAAAAGAGCAAGAAAAGAAGAAGTCTAAGAGCAAGAAAGCTTCAAAGATAAA
 GAAAAGAAATCGAAAGACAAGCAAGCAGCTCTGACGATGAAAATGAGACAAAAAAGG
 TTTCGATTTTGCTTCAGAAGATGGTATTATTCTACACCAATCCCAAATATGATGGTGCC
 AATATTATTCCGAAGCAAACCGAGATTGCTCAAATCTATCCTGATATTCAAACAAAG
 AAAAGTGTAAATCACCTATTATGCTCTGATGATGTTGTTCTATGAAAAAGGGGC
 AAACCGCTCGTCTTCCTGGAAAAAGGGAAATGACAAGGTTGTTATTGAAGGAAA
 ATTAACAATGTCGCTTCATCAGCAACTACTAAAAAGGAATCTCTTAAGGTTAC
 TGCCAAAGTAAAGGTTCTAAGAAAATAGCAAACCTCATCAAGTATGGTATGACAGGCA
 AGACAGTCACTGTCATTGATAAAAAGACTTATTTGATTATTCAAAGATAAATTACTG
 CATAAAATGGATAAT

(D.)

S. mutans ComB protein

MDPKFLQSAEFYRRRYHNATLLIVPLVCLIIFLVIFLCFAKKEITVISTGEVAPTKVV
 DVIQSYSDSSIKNNLDDNAAVEKGDVLIYESENASPNRQTEQKNIKERQKREEEKK
 KHQKSKKKKSKSKASKDKKKKSDKESSSDDENETKKVSIFASEDGIIHTNPKYDGA
 NIIPKQTEIAQIYPDFIQKTRKVLITYYASSDDVVSMMKGQTARLSLEKKGNDKVVIEGK
 INNVASSATTKKGNLFKVTAKVKVSKKNSKLICKYGMTGKTVTVIDKKTYFDYFKDKLL
 HKMDN

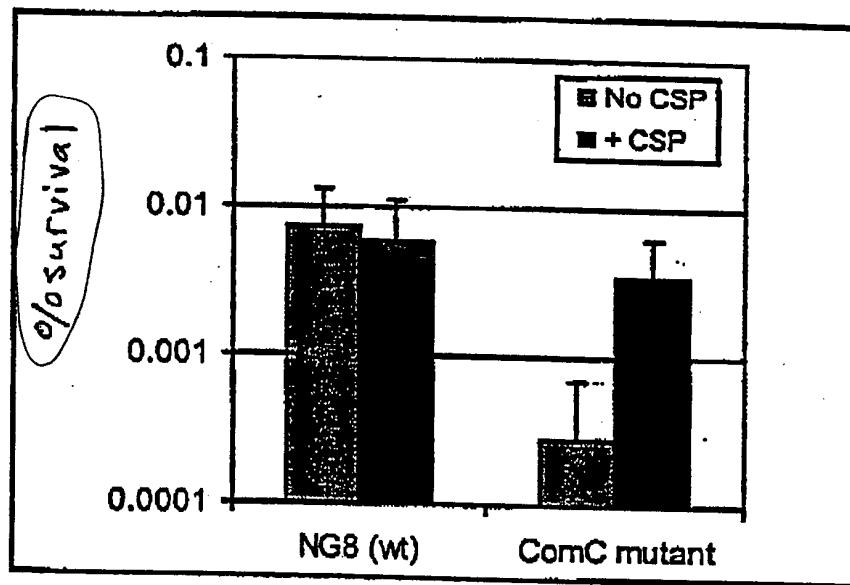


Figure 12